Interphase ribosomal RNA cistron staining in thyroid epithelial cells in Grave’s disease, Hashimoto’s thyroiditis and benign and malignant tumours of the thyroid gland

N N Mamaev, E N Grynyeva, Y V Blagosklonnaya

Abstract

Aim—To evaluate the expression of ribosomal cistrons in human thyroid epithelial cells (TECs) of patients with Grave’s disease, Hashimoto’s thyroiditis and benign and malignant tumours of the thyroid gland.

Methods—TEC nucleoli were investigated in fine needle biopsy specimens from 10 controls, 39 patients with Grave’s disease, 15 with Hashimoto’s thyroiditis, 56 with benign, and 15 with malignant tumours of the thyroid. A one step silver staining method was applied. In most cases serum concentrations of thyroxine and triiodothyronine as well as goitre size were determined. In every case 100 TECs were evaluated for the mean numbers of nucleoli and for the average number of argyrophilic nucleolar organiser regions (AgNORs) per nucleus.

Results—NORs were activated in all patients, but not in controls. The numbers of AgNORs in patients with Grave’s disease were closely correlated with thyroxine or triiodothyronine, or both, concentrations and with the size of the thyroid. In patients with Hashimoto’s thyroiditis about 30% of TECs nucleoli did not contain AgNORs, whereas others were heavily impregnated with silver. Compared with controls and benign tumours, the nucleoli of carcinomatous TECs were larger and irregular in shape. The mean number of AgNORs per nucleus in malignant cells was higher than that in their benign counterparts.

Conclusions—The mechanism by which NORs are activated in TECs varies depending on the type of lesion. The higher AgNOR score in TECs from malignant tumours can be used to distinguish them from their benign counterparts.

Keywords: thyroid disease, thyroid epithelial cells, nucleolar organiser regions, silver staining.

The method of argyrophilic nucleolar organiser region (AgNOR) staining has been widely described. This technique permits the evaluation of pre-ribosomal RNA transcription by means of light microscopy. As RNA transcription, ribosome formation and protein synthesis are essential, not only for cell proliferation and differentiation but also for general functional status, AgNOR counting is important in the study of many human cell types. Furthermore, as thyroid epithelial cells (TECs) are both hormone producing and hormone dependent cells, they may be suitable subjects for the study of metabolic variation. There are, however, few published papers describing AgNOR behaviour in human TECs. Most studies have been carried out on cells isolated from patients with thyroid tumours. The aim of this paper was to compare the numbers of nucleoli and AgNORs in TECs in healthy subjects and patients with histologically confirmed thyroid disease (Grave’s disease, Hashimoto’s thyroiditis, and benign and malignant tumours of the thyroid).

Methods

Thyroid specimens from 29 women and 10 men with Grave’s disease, 15 women with Hashimoto’s thyroiditis, and 56 patients with benign (all women) and 15 with malignant (three men and 12 women) tumours of the thyroid were studied. In most cases serum concentrations of thyroxine and triiodothyronine, as well as goitre size (graded as recommended by the World Health Organisation (WHO)), were measured.

Twenty six patients with Grave’s disease were hyperthyroid. Their diagnosis was based on the typical clinical picture, elevated serum thyroxine and triiodothyronine concentrations, and increased 131I uptake. Clinical examination revealed a soft diffuse goitre in all of the patients and infiltrative ophthalmopathy in 50%. Other clinical features, such as fatigue, loss of weight, increased appetite, tachycardia, and palpitations, were also noted. Mean (SEM) serum thyroxine and triiodothyronine concentrations, measured by radioimmunoassay, were 248 (33.9) nmol/l (normal range 60–120 nmol/l) and 3.78 (0.32) nmol/l (normal range 1.17–2.18 nmol/l), respectively.

Thirteen patients with Grave’s disease were euthyroid, mainly because they had been taking methimazole for five to 36 months before the study was undertaken. Mean (SEM) serum thyroxine and triiodothyronine concentrations in this group were 1311 (72.4) and 1.5 (0.1) nmol/l, respectively. Goitre sizes were as follows: grade I in one patient; grade II in 23; and grade III in 16.
Interphase ribosomal RNA cistron staining of TELs in thyroid disease

Table 1 Results of silver staining of TECs isolated from controls and patients with thyroid disease. Values are expressed as mean (SEM)

<table>
<thead>
<tr>
<th>Diagnosis</th>
<th>Patients (n)</th>
<th>Nucleoli (n)</th>
<th>AgNORs (n)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>12</td>
<td>1.61 (0.07)</td>
<td>5.36 (0.36)</td>
</tr>
<tr>
<td>Grave’s disease</td>
<td>26</td>
<td>3.52 (0.09)*</td>
<td>9.82 (0.24)†</td>
</tr>
<tr>
<td>Hyperthyroid</td>
<td>13</td>
<td>3.24 (0.22)*</td>
<td>8.34 (0.4)*</td>
</tr>
<tr>
<td>Hashimoto’s thyroiditis</td>
<td>15</td>
<td>2.90 (0.14)*</td>
<td>7.87 (0.3)*</td>
</tr>
<tr>
<td>TEC</td>
<td>15</td>
<td>2.56 (0.16)</td>
<td>14.25 (1.56)</td>
</tr>
<tr>
<td>Lymphocyte</td>
<td>15</td>
<td>2.46 (0.11)*</td>
<td>8.01 (0.12)†</td>
</tr>
<tr>
<td>Colloidal goitre</td>
<td>24</td>
<td>2.47 (0.11)*</td>
<td>7.95 (0.2)</td>
</tr>
<tr>
<td>Follicular adenoma</td>
<td>32</td>
<td>2.55 (0.22)*</td>
<td>14.1 (0.78)†</td>
</tr>
</tbody>
</table>

*p < 0.05, tp < 0.005 v controls; n = number.

All of the patients with Hashimoto’s thyroiditis had firm, longstanding goitre, persisting for six months to 30 years (mean 11.1 years). Goiter sizes were grade II in nine patients and grade III in six. Fourteen patients were euthyroid and only one presented with hyperthyroidism. Thyroglobulin antibodies were detected in 10 patients. Titres of 1:320 and higher were regarded as significant and these were found in seven patients. Diagnosis of Hashimoto’s thyroiditis was confirmed by fine needle aspiration cytology.

Controls were obtained from two sources. Normal thyroid glands were removed at necropsy from seven patients who had died of ischaemic heart disease. Specimens were also obtained from five volunteers with normal thyroid glands.

Fine needle biopsy specimens of the thyroid were air dried, fixed in 1:3 acetic acid: methanol for 10 minutes and impregnated with silver as described by Howell and Black13 with a slight modification14—that is, pretreatment in 2 N formic acid solution for 10 minutes. Pretreating the specimens with formic acid helps to reduce loss of silver from the nucleoli, making evaluation of the numbers, sizes, and patterns of nucleoli, as well as AgNOR counting, much easier. The preparations were then rinsed in distilled water and lightly counterstained with 2% Giemsa phosphate buffer (pH 6.8) for three seconds.

The numbers of nucleoli and of individual AgNOR dots in each cell were counted at a magnification of x1250. At least 100 interphase cells, chosen at random from each sample, were examined. When AgNORs occurred in large clumps, they were counted approximately, using the area of one typical dot as a standard for determining the possible numbers of AgNORs in each clump. The mean numbers of nucleoli and AgNORs in each nucleus were calculated. The Student’s t test was used to compare data in the patient and control groups.

Results

controls

The results of AgNOR staining are presented in figs 1A and 2. Each TEC nucleus contained a mean of 1.6 (range 1.34–1.96) nucleoli. The number of AgNORs in nucleoli, corresponding to TEC pre-ribosomal RNA transcription and ribosome maturation, ranged from 1 to 15 per nucleus. The mean numbers of AgNORs in nucleoli ranged from 3.0 to 6.9 (mean 5.36) per nucleus.

Grave’s disease

TEC nucleoli in patients with Grave’s disease were larger than those in controls (fig 1B). Their numbers ranged from one to seven (table 1), significantly higher than in controls. Further analysis of silver stained TECs in the

Figure 1 Representative photomicrographs of silver stained TECs in (A) a normal subject, and in patients with (B) Grave’s disease, (C) follicular adenoma, and (D) cancer. (Original magnification, x1200).
groups of patients with and without hyperthyroidism revealed that the mean numbers of nucleoli in these cells ranged from 2.5 to 4.3 and from 2.3 to 4.06 (mean 3.5 and 3.24), respectively.

The AgNOR score of TECs in patients with Grave’s disease was also higher (figs 1B and 2) than in controls and was strongly related to peripheral blood concentrations of thyroxine and triiodothyronine (fig 3). Furthermore, in euthyroid patients a direct association was observed between the number of AgNORs in TEC nuclei and the size of the thyroid (fig 4). Thus, TEC nucleolar activity in patients with grade III goitre was higher than in those with grades I or II; mean (SEM) 9.26 (0.64) compared with 7.55 (0.24) AgNORs per nucleus (p < 0.05).

**Hashimoto’s Thyroiditis**

The nucleoli of TECs from patients with Hashimoto’s thyroiditis were smaller than those from controls and patients with Grave’s disease. Their numbers ranged from one to seven (table 1), although occasional TECs contained no nucleoli at all. The mean number of nucleoli in TECs from patients with Hashimoto’s thyroiditis ranged from 2.1 to 4.0 per nucleus (mean 2.9). These values were higher than those in controls (p < 0.0005), but lower than those in patients with Grave’s disease with and without hyperthyroidism.

The numbers of AgNORs in TEC nuclei from patients with Hashimoto’s thyroiditis ranged from one to 80 per nucleus; 1–30% of TECs had no AgNORs. The number of TECs ranged from 5.5 to 10.2 per nucleus (mean 7.87), significantly higher (p < 0.001) than that in controls but significantly lower (p < 0.001) than that in patients with Grave’s disease with (mean 9.82) or without (mean 8.34) hyperthyroidism. However, when the numbers of AgNORs were plotted per nucleolus, very similar histogram patterns were produced for patients with Hashimoto’s thyroiditis and Grave’s disease (fig 5). Interestingly, the highest scores were seen in patients with Hashimoto’s thyroiditis and symptomatic hyperthyroidism. Conversely, the lowest scores were found in patients with Hashimoto’s thyroiditis without hyperthyroidism.

The AgNOR score was also raised in lymphocytes isolated from patients with Hashimoto’s thyroiditis (table 1) (mean 11.25 AgNORs per nucleus). This is presumably because of damage to TECs, resulting in lymphocyte activation.

**Benign Tumours**

The results presented in fig 1C and table 1 show that the mean (SEM) numbers of nucleoli in TECs in patients with nodular goitres (2.5 (0.11) and follicular adenomas (2.5 (0.11)) were higher than those in controls (1.2 (0.07); p < 0.05). There was also a statistically significant difference (p < 0.005) in TEC AgNOR counts in these three groups (8.0 (0.12), 7.95 (0.2) and 5.36 (0.36) AgNORs per nucleus).
AgNOR scores were moderately increased in patients with benign tumours and Hashimoto's thyroiditis.

The proliferative activity of TECs isolated from patients with Grave's disease is minimal, suggesting that the increase in ribosomal cistron activity may result primarily from TEC hormonal activity and hyperplasia.

The size and argyrophilia of TEC nuclei in patients with Hashimoto's thyroiditis were more heterogenous than in patients with Grave's disease and controls. Thus about 30% of TECs from patients with Hashimoto's thyroiditis did not contain AgNORs, whereas others were heavily impregnated with silver. As a result, the mean (SEM) number of AgNORs in TECs from patients with Hashimoto's thyroiditis, containing at least one silver stained nucleolus, was higher (7.9 (0.3); p < 0.001) than that in controls (5.4 (0.36)) but significantly lower than that in TECs from patients with Grave's disease with and without hyperthyroidism (9.82 (0.24) and 8.34 (0.4), respectively; p < 0.001). It is worth noting that there was no association between AgNOR scores and the size of the thyroid in patients with Hashimoto's thyroiditis. These findings are in agreement with previous reports that enlargement of the thyroid in patients with Hashimoto's thyroiditis is more closely related to lymphoid infiltration than to TEC hyperplasia.

The mean number of nuclei in TECs from patients with thyroid tumours did not differ significantly from that in controls, whereas the mean number of AgNORs per nucleus was greatly increased in the former. In our opinion, this increase in ribosomal cistron activity in tumour cells may be explained by partial activation of their proliferative potential, which in turn might be stimulated either by oncogenes or by other biologically active substances.

In conclusion, our findings demonstrate that the silver staining method can be used to assess the functional status of human TECs in both physiological and pathological conditions. We believe that AgNOR counting in TECs may prove to be very useful in distinguishing malignant from benign lesions of the thyroid.