Concomitant progressive multifocal leucoencephalopathy and primary central nervous system lymphoma expressing JC virus oncogenic protein, large T antigen

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Abstract
This report describes the concomitant occurrence of the JC virus (JCV) induced demyelinating disease progressive multifocal leucoencephalopathy (PML) and a primary central nervous system lymphoma (PCNS-L) in a patient with AIDS. Postmortem neuropathological examination revealed characteristic features of PML including multiple lesions of demyelination, enlarged oligodendrocytes with hyperchromatic nuclei (many containing eosinophilic intranuclear inclusions), and enlarged astrocytes with bizarre hyperchromatic nuclei. Immunohistochemical analysis demonstrated the expression of the JCV capsid protein VP-1 in the nuclei of infected oligodendrocytes and astrocytes. The PCNS-L lesion located in the basal ganglia was highly cellular, distributed perivascularly, and consisted of large atypical plasmacytoid lymphocytes. Immunohistochemical examination of this neoplasm identified it to be of B cell origin. Moreover, expression of the JCV oncogenic protein, T antigen, was detected in the nuclei of the neoplastic lymphocytes. This study provides the first evidence for a possible association between JCV and PCNS-L.

Keywords: JC virus; progressive multifocal leucoencephalopathy; primary central nervous system lymphoma; acquired immunodeficiency syndrome; demyelination; T antigen

Progressive multifocal leucoencephalopathy (PML) is a human demyelinating disease that results from the selective destruction of myelin producing oligodendrocytes by the human polyoma JC virus (JCV) (reviewed by Major et al.). Seroepidemiological studies indicate that JC virus infection is very common, with approximately 70–90% of adults possessing JCV specific antibodies. Despite this high rate of infection, PML usually emerges as an opportunistic infection of the central nervous system (CNS) in immunocompromised patients. Although once considered a relatively rare disease, PML has become more common as a result of AIDS (reviewed in Berger and Concha), and is estimated to occur in up to 5% of all patients with AIDS. In addition to CNS opportunistic infections such as PML, CNS neoplastic diseases occur with increased frequency in immunosuppressed patients, including primary CNS lymphoma (PCNS-L). Although PML and PCNS-L are common in immunosuppressed patients, their concomitant occurrence is relatively rare. Here, we describe the rare co-occurrence of PML and PCNS-L and demonstrate expression of JCV large tumour antigen (T antigen) in the tumour cells. In the light of earlier observations of the presence of JCV in B cells and the oncogenic potential of the JCV early protein, we discuss the importance of these findings in the pathogenesis of PCNS-L.

Case report
A 33 year old homosexual man with a five year history of AIDS was admitted to Thomas Jefferson University Hospital, Philadelphia after being found with altered mentation in his parked car. AIDS had been complicated by three episodes of Pneumocystis carinii pneumonia (PCP), gastroparesis, pancreatitis related to pentamidine, herpes, and candida oesophagitis. The patient completed a course of antibiotics for pseudomonas septicaemia six weeks before admission.

On admission, the patient was awake but disoriented to time and place and unable to follow commands. His temperature was 101.5°F, blood pressure was 115/70 mm Hg, heart rate was 84 beats/minute, and respiration was 16 breaths/minute and unlaboured. The patient was not able to cooperate fully with the neurological examination, but cranial nerves II–XII were intact and the deep tendon reflexes were symmetric and 3+ in the upper extremities and 2+ in the lower extremities. Gait was unsteady. On admission, complete blood cell count revealed a white blood cell count of 1.2×10^9/litre, haemoglobin was 86 g/litre, haematocrit 24.6, and platelets were 134×10^3/litre. Cerebrospinal fluid (CSF) was clear and colourless, with a normal glucose of 500 mg/litre, haemotocrit 24.6, and platelets were 134×10^3/litre. Cerebrospinal fluid (CSF) was clear and colourless, with a normal glucose of 500 mg/litre (normal, 400–700), and a slightly increased protein concentration of 1010 mg/litre (normal, 150–500). Gram stain, acid fast stain, and India ink preparation of the CSF revealed no microorganisms, and bacterial antigens were negative for Haemophilus influenzae, Neisseria meningitidis, and Streptococcus pneumoniae. Toxoplasmosis titre, serum cryptococcal antigen, and blood cultures for bacteria and Mycobacterium avium intracellulare (MAI) were negative. Cytomegalovirus titre was positive one week before admission.
Axial computed tomography (CT) images of the brain were obtained on admission and compared with images obtained three months previously. CT scan showed new bilateral, inferior frontal white matter hypoattenuation with no enhancement, stable cortical and central atrophy, and worsening sphenoid sinus inflammation. Chest radiographs were within normal limits. Magnetic resonance imaging with gadolinium two days after admission revealed atrophy, prominence of the right temporal horn, and abnormal T2 weighted images in the frontal, temporal, and parietal lobes (fig 1A).

Soon after admission the patient experienced a tonic clonic seizure and was treated with valium and dilantin without subsequent seizure activity. The patient received an empirical 11 day course of intravenous vancomycin, mezlocillin, and gentamycin. Chest radiographs five days after admission revealed upper lobe infiltrates and clindamycin and primaquin were administered at therapeutic doses to treat possible PCP (because the patient had an allergy to sulphur and a history of pancreatitis related to pentamidine). Although his mental status improved 10 days after admission, it worsened progressively over the next weeks. Twelve days after admission, ciprofloxacin and clarithromycin were started as empirical treatment for MAI because of a persisting temperature increase to 105°F. The patient developed swallowing difficulty and eventually was unable to take anything by mouth. Parenteral morphine was administered as needed to maintain comfort and the patient died 29 days after admission.

**Postmortem examination**

Postmortem examination of the brain revealed multiple diffuse patches of discolouration and softening of the subcortical white matter in the frontal lobe, especially towards the ventral surface of the brain, where they had a cavitary appearance (fig 1B). This destructive process extended caudally into the right parietal and temporal lobes, below the right putamen, and into the occipital lobe.

Histological sections of these lytic areas showed multiple, small and moderate sized areas of demyelination, some of them confluent, with relative good preservation of axons (fig 2A, B). In the areas of demyelination, there were multiple enlarged oligodendrocytes, many containing eosinophilic intranuclear inclusions, and numerous enlarged bizarre astrocytes with pleomorphic cytoplasm and hyperchromatic nuclei (fig 2C, D). Immunohistochemical analysis demonstrated the expression of the viral capsid protein VP-1 in the nuclei of oligodendrocytes and transformed astrocytes (fig 2E, F). In the cortex of the frontal and temporal lobes, small collections of reactive lymphocytes, plasma cells, and neutrophils consistent with microabscesses were present.

Sections from the basal ganglia revealed a highly cellular neoplasm, characterised by a homogenous population of small round cells with pleomorphic nuclei, which diffusely infiltrated the surrounding tissue (fig 3A, B). Some mitotic figures were present. There were also small necrotic patches within the neoplasm. In the periphery of the tumour, the neoplastic cells had a perivascular concentric distribution and formed collars around blood vessels. Immunohistochemical examination using monoclonal antibodies specific for B cell surface antigens CD20 and CD79a showed...
that this neoplasm was of B cell origin (fig 3C, D). According to the Working Formulation and latest World Health Organisation classification of tumours of the CNS, this neoplasm was classed as a diffuse large cell immunoblastic lymphoma. Because numerous studies have detected JCV within B cells (reviewed by Gallia et al), we next investigated the expression of JCV in the PCNS-L. As shown in fig 3E, F, JCV T antigen was expressed in the nuclei of the neoplastic lymphocytes, with approximately 60% of the tumour cells being immunopositive for JCV T antigen.

Other notable pathological findings included AIDS cardiomyopathy, focal pulmonary fibrosis, candida septicaemia with multiple abscesses in the kidneys, heart, lungs, and brain, and secondary testicular atrophy, Sertoli cell only.

Discussion

CNS dysfunction is a prominent feature in patients with AIDS. Involvement of the CNS occurs clinically in approximately 30–40% of patients with AIDS, and postmortem examination reveals neuropathological abnormalities in up to 80–90% of patients. In addition to neurological disease caused by direct infection, the CNS of individuals infected with the human immunodeficiency virus type 1 (HIV-1) is susceptible to a unique constellation of opportunistic infections and neoplasms.
Moreover, multiple CNS infections and neoplasms are reported to occur in between 10% and 15% of patients with AIDS.\textsuperscript{16} We describe a patient with AIDS and concomitant PML and PCNS-L. Moreover, we demonstrate the expression of the early viral protein, T antigen, within the nuclei of the neoplastic lymphocytes.

The concomitant occurrence of PML and PCNS-L in patients with and without AIDS is a relatively rare event. Table 1 summarises the reported co-occurrences of these two CNS diseases. There are several reports describing the co-occurrence of these two diseases in patients without AIDS.\textsuperscript{21a,22–25} Davies \textit{et al} reported a patient with PML and a small granulomatous lesion consisting of lymphocytes, plasma cells, histiocytes, and large binucleate cells in the frontal white matter.\textsuperscript{22} A similar cellular infiltrate was present around many intracerebral blood vessels and leptomeninges. The authors considered these lesions as possible evidence of Hodgkin’s disease or other reticulosis limited to the CNS. The description of the granulomatous lesion is consistent with PCNS-L. GiaRusso and Koeppen\textsuperscript{24} described a small PCNS-L in a patient with atypical progressive multifocal leukoencephalopathy. The PCNS-L in this case was thought to arise directly within the foci of demyelination. Concomitant PML and PCNS-L have also been reported in patients treated with immunosuppressive agents after renal transplantation.\textsuperscript{22,23} In addition to these non-AIDS associated

\begin{figure}[h]
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\caption{Histological and immunohistochemical evaluation of primary central nervous system lymphoma. Neoplastic cells have an angiocentric pattern, forming collars around blood vessels (A; original magnification, $\times20$; haematoxylin and eosin). At higher magnification the neoplastic cells are round with big nuclei and moderate eosinophilic cytoplasm (B; original magnification, $\times100$; haematoxylin and eosin). Immunohistochemical staining with anti-CD20 and anti-CD79a antibodies shows intense cytoplasmic and membrane associated reactivity in the neoplastic perivascular cells (C; original magnification, $\times20$; D, original magnification, $\times100$). Immunohistochemical staining with an antibody detecting JC virus T antigen shows expression in the nuclei of approximately 60% of the neoplastic lymphocytes (E; original magnification, $\times40$; F; original magnification, $\times100$).}
\end{figure}
Table 1  Association of PML and primary CNS lymphomas

<table>
<thead>
<tr>
<th>Patient</th>
<th>Underlying disease</th>
<th>CNS lymphoma reported</th>
<th>Other neuropathological features</th>
<th>Ref</th>
</tr>
</thead>
<tbody>
<tr>
<td>33 M</td>
<td>AIDS</td>
<td>B cell immunoblastic lymphoma</td>
<td>PML</td>
<td>Our study</td>
</tr>
<tr>
<td>44 M</td>
<td>AIDS</td>
<td>Large cell lymphoma</td>
<td>HIV related encephalopathy</td>
<td>Morgello (1992)</td>
</tr>
<tr>
<td>68 F</td>
<td>Immunosuppression secondary to renal transplant</td>
<td>Histiocytic lymphoma</td>
<td>PML</td>
<td>Ho et al (1980)</td>
</tr>
<tr>
<td>82 M</td>
<td>CLL</td>
<td>Immunoblastic sarcoma (reticulum cell sarcoma)</td>
<td>PML (atypical)</td>
<td>GiaRusso and Koeppen (1978)</td>
</tr>
<tr>
<td>41 F</td>
<td>NR</td>
<td>Hodgkin's disease</td>
<td>PML</td>
<td>Davies et al (1973)</td>
</tr>
</tbody>
</table>

CLL, chronic lymphocytic leukemia; CNS, central nervous system; F, female; HIV, human immunodeficiency virus; M, male; PML, progressive multifocal leukoencephalopathy; NR, not reported; CMV, cytomegalovirus.

 Epstein-Barr virus (EBV) has been implicated in the pathogenesis of AIDS related PCNS-L, with between 50% and 100% of AIDS related CNS lymphomas containing various EBV sequences. The existence of EBV negative PCNS-L suggests that other factors may be involved in the pathogenesis of at least some AIDS related PCNS-L cases and, recently, human herpesvirus 8 (HHV-8) has been detected in CNS lymphomas from patients with and without AIDS. Together, these observations raise the possibility that several viruses may contribute to the development of PCNS-L.

JCV may indirectly influence the development of PCNS-L. Morgello et al reported an increased incidence of viral CNS infections in patients with AIDS and CNS lymphomas, suggesting that viral infection might play a role in the pathogenesis of CNS lymphomas in the immunocompromised individual. Although PML was only detected in one of 15 patients with primary CNS lymphoma, and that patient also had CMV encephalitis, perhaps several CNS viral infections can provide an initiating event for the development of PCNS-L in the immunocompromised patient.

In this report, we describe the co-occurrence of PML and PCNS-L in a patient with AIDS and demonstrate the presence of T antigen within the nuclei of a primary CNS B cell lymphoma. We thank Dr B Fenderson for assistance in retrieving some of the patient records, S Enam for assistance with immunohistochemistry, Drs S Morgello and J Martinez for helpful suggestions, and Ms C Schriver for editorial assistance. This work was supported by grants from the NIH awarded to KK. Dr G Gallia’s current address is: Department of Neurosurgery, John Hopkin’s University Hospital, Baltimore, USA.