Morphoea and *Borrelia burgdorferi*: results from the Scottish Highlands in the context of the world literature

J R Goodlad, M M Davidson, P Gordon, R Billington, D O Ho-Yen

J Clin Pathol: Mol Pathol 2002;**55**:374–378

See end of article for authors' affiliations

Correspondence to:
Dr J R Goodlad,
Department of Pathology,
Highland Acute Hospitals
NHS Trust, Raigmore
Hospital, Old Perth Road,
Inverness IV2 3UJ, UK;
John.Goodlad@
raigmore.scot.nhs.uk

Accepted for publication
11 September 2002

Erythema chronicum migrans, acrodermatitis chronica atrophicans (ACA), and lymphocytoma cutis are all cutaneous manifestations of Lyme disease, a multisystem disorder that follows infection with *Borrelia burgdorferi*.^{1–4} An aetiological role for *B burgdorferi* has also been proposed for other skin disorders, including primary cutaneous B cell lymphoma (PCBCL) and morphoea. Although there is increasing evidence to support this hypothesis in the context of PCBCL,⁵ studies investigating the link between *B burgdorferi* and morphoea have produced conflicting results, some reports suggesting a positive association, but others not.⁶ This has led to the proposal that regional variations in *B burgdorferi* may be important in dictating the spectrum of clinical disease following infection, with morphoea only being caused by certain subspecies endemic to specific geographical areas.⁷

"Studies investigating the link between *Borrelia burgdorferi* and morphoea have produced conflicting results, some reports suggesting a positive association, but others not"

Although Lyme disease is endemic in many areas of the UK, only two previous studies have investigated the possible link with morphoea specifically in the UK.^{8,9} Both gave a negative result, but this was based primarily on patients with morphoea lacking serological evidence of previous infection with *B burgdorferi*. However, because *B burgdorferi* infection may occur without the production of specific antibodies these results are inconclusive.¹⁰ Therefore, we used a polymerase chain reaction (PCR) technique to evaluate skin biopsies taken from patients with morphoea in the Highlands of Scotland, an area with endemic Lyme disease, for the presence or absence of *B burgdorferi* specific DNA. Recently, this approach has been used successfully to demonstrate a significant

Aims: Previous studies investigating the link between infection with *Borrelia burgdorferi* and morphoea have produced conflicting results. Often, these studies have been undertaken in patients from different regions or countries, and using methods of varying sensitivity for detecting *Borrelia burgdorferi* infection. This study aimed to establish whether a relation could be demonstrated in the Highlands of Scotland, an area with endemic Lyme disease, with the use of a sensitive method for detecting the organism.

Methods: The study was performed on biopsies of lesional skin taken from 16 patients from the Highlands of Scotland with typical clinical features of morphoea. After histological confirmation of the diagnosis, a nested polymerase chain reaction (PCR) using primers to a unique conserved region of the *Borrelia burgdorferi* flagellin gene was performed on DNA extracts from each biopsy. A literature search was also performed for comparable studies.

Results: None of the 16 patients had documented clinical evidence of previous infection with *B burgdorferi*. DNA was successfully extracted from 14 of the 16 cases but all of these were negative using PCR for *B burgdorferi* specific DNA, despite successful amplification of appropriate positive controls in every test. The results were compared with those of other documented studies.

Conclusions: Examination of the literature suggests that there is a strong geographical relation between *B burgdorferi* and morphoea. These results, in which no such association was found, indicate that morphoea may not be associated with the subspecies of *B burgdorferi* found in the Highlands of Scotland.

association between *B burgdorferi* and PCBCL in patients from the same region.⁵ A literature review was also performed to interpret our results in the context of the possible geographical variations that may exist in the relation between *B burgdorferi* infection and the subsequent development of morphoea.

MATERIALS AND METHODS

Case selection

The pathology department, Raigmore Hospital, Inverness, UK, processes all surgical pathology specimens for the Highland and Western Isles regions of Scotland, an area with a population of approximately 250 000 in which Lyme disease is endemic. By searching the surgical pathology files, Raigmore Hospital, Inverness (from 1976 to date), and reviewing case records of all patients with histology compatible with morphoea we were able to identify 16 patients for study, all with histological and clinical features typical of morphoea.

Pathological studies

Sections (5 µm thick) were cut from formalin fixed, routinely processed, paraffin wax embedded tissue blocks of each biopsy and stained with haematoxylin and eosin. Each case was assessed to confirm a diagnosis of morphoea and to approximate the stage of the disease using standard criteria.¹¹

Demonstration of *B burgdorferi* DNA

Sections (6 × 5 µm) cut from formalin fixed, paraffin wax embedded tissue blocks were dewaxed and DNA extracted using the Intergen EX-WAX DNA extraction kit (Intergen,

Abbreviations: ACA, acrodermatitis chronica atrophicans; PCBCL, primary cutaneous B cell lymphoma; PCR, polymerase chain reaction

Figure 1 Haematoxylin and eosin stained section (original magnification, $\times 20$) showing early changes of morphoea. A perivascular infiltrate is present in the dermis and the superficial subcutis. The reticular dermis appears pale owing to the presence of swollen collagen bundles.

Figure 2 Haematoxylin and eosin stained section (original magnification, $\times 100$) showing later stage morphoea. Thickened collagen bundles fill the reticular dermis in the vicinity of an eccrine gland that has lost the normal surround of adipose tissue.

Manhattanville, New Jersey, USA). All DNA extracts were tested by PCR with primers for β globin to ensure that DNA of sufficient quality and quantity was obtained for subsequent analysis.

To detect *B burgdorferi*, a nested PCR assay was used to amplify a sequence of the highly conserved flagellin encoding region, as described previously.¹² The first stage PCR was performed with 20 μ l of sample in a reaction volume of 50 μ l, containing final concentrations of 10mM Tris/HCl (pH 8.5), 50mM KCl, 3.5mM MgCl₂ (Applied Biotechnologies), 0.2mM dNTP (Pharmacia Biosystems, Milton Keynes, UK), 0.5 U Taq polymerase (Applied Biotechnologies), and 0.5 μ M of each primer F1 (5'-ATT AAC GCT GCT AAT CTT AGT-3') and F3 (5'-GTA CTA TTC TTT ATA GAT TC-3') (Seyvern Biotech, Kidderminster, UK). The thermal cycling conditions were 35 cycles of one minute at 94°C, two minutes at 41°C, and three minutes at 66°C, followed by a further extension period of five minutes at 72°C. For the nested PCR, 20 μ l of a 1/5 dilution of the first stage product was amplified in a 50 μ l reaction mix with a reduced concentration of MgCl₂ (2.5mM) and primers F6 (5'-TTC AGG GTC TCA AGC GTC TTG GAC T-3') and F8 (5'-GCA TTT TCA ATT TTA GCA AGT GAT G-3') (Seyvern Biotech). The thermal cycling conditions were 35 cycles of one minute at 94°C, one minute at 50°C, and one minute at 72°C, with a final extension of five minutes at 72°C. A positive control (~ 10 cultured *B burgdorferi* organisms) and negative control were included in all cases. The PCR products were visualised under ultraviolet transmission of ethidium bromide stained agarose gels.

Figure 3 Agarose gel showing results after the polymerase chain reaction with *Borrelia burgdorferi* flagellin gene. Lanes 1 and 6, DNA molecular weight markers; lane 2, positive control; lane 3, negative control; lanes 4 and 5, negative results from DNA extracts of two cases of morphoea.

Literature review

A textword Medline search was conducted for the years 1966 to date using the terms “morphea”, “morphoea”, “Lyme disease”, and “Borrelia” in various combinations. Abstracts of all articles were read and papers selected for inclusion if they were published in English and used laboratory techniques to investigate the relation between *B burgdorferi* infection and morphoea.

RESULTS

Clinical features

There were five male and 11 female patients ranging in age from 3 to 69 years at diagnosis. According to the classification of Peterson *et al*,¹³ there were 13 cases of plaque morphoea (12 morphoea en plaque, one atrophoderma of Pasini and Pierini), two cases of linear morphoea (one linear morphoea, one en coup de sabre), and one case of generalised morphoea. None of the patients had a history of antibiotic treatment in their hospital notes and none had a documented history of manifestations of Lyme disease.

Pathological findings

All 16 cases showed histological features consistent with a diagnosis of morphoea, the principle finding being the presence of thickened collagen bundles in the reticular dermis. The histological stage of the lesion was approximated by additional changes.¹¹ Three cases were interpreted as early stage lesions. All displayed a moderate lymphocytic infiltrate with fewer numbers of plasma cells arranged around blood vessels lined by swollen endothelial cells (fig 1). Six biopsies were regarded as showing features of late stage lesions. In these, there was increased eosinophilia of the thickened collagen bundles, together with homogeneous collagen in the papillary dermis, atrophy of eccrine glands with marked loss of periglandular fat, some fibrous thickening of blood vessel walls, and a minimal or absent chronic inflammatory cell infiltrate (fig 2). The remaining seven cases displayed features in keeping with lesions of an intermediate duration in which the “late” changes were either less pronounced, or only some were present.

PCR analysis

Amplifiable DNA was obtained from 14 of the 16 biopsies, as determined by PCR with primers for β globin. These 14 cases were tested using primers for the *B burgdorferi* flagellin gene. In each test, there was successful amplification of control DNA extracted from cultured borrelia organisms, giving a product of 275 bp after the nested run. However, all 14 samples from patients with morphoea were negative, as were the negative controls (fig 3).

Table 1 Summary of previous studies investigating the relation between *Borrelia burgdorferi* infection and morphoea in different geographical locations

Reference	Country	Direct visualisation in tissue sections of morphoea biopsies	Serology	Culture from biopsies of morphoeic lesions	PCR analysis of DNA extracts of morphoea biopsies
Aberer <i>et al</i> (1987) ¹⁴	Austria	3/18 (immunoperoxidase)	8/15	1/4	ND
Aberer <i>et al</i> (1988) ¹⁵	Austria	3/9 (immunoperoxidase)	5/9	ND	ND
Aberer <i>et al</i> (1991) ¹⁶	Austria	ND	14/30	1/11	ND
Aberer and Stanek (1987) ¹⁷	Austria	4/13 (immunoperoxidase)	7/13	1/3	ND
Breier <i>et al</i> (1996) ¹⁸	Austria	ND	13/39*	ND	ND
Aberer <i>et al</i> (1996) ¹⁹	Austria	4/16 (immunoperoxidase)	ND	0/22	ND
Breier <i>et al</i> (1999) ²⁰	Austria	0/1 (silver stain)	1/1	1/1†	ND
Fujiwara <i>et al</i> (1997) [‡]	Germany	ND	ND	ND	3/4
Schempp <i>et al</i> (1993) ²¹	Germany	ND	0/5	ND	9/9
Wienecke <i>et al</i> (1995) ²²	Germany	ND	4/18	ND	0/30
Weide <i>et al</i> (2000) ²³	Germany	ND	3/53	ND	0/33
Weber <i>et al</i> (1988) ²⁴	Germany	ND	0/2	2/2	ND
Muhlemann <i>et al</i> (1986) ⁸	UK	ND	0/22	ND	ND
Lupoli <i>et al</i> (1991) ⁹	UK	ND	0/31	0/9	ND
Raguin <i>et al</i> (1992) ²⁵	France	0/14 (immunoperoxidase)	0/15	0/10	ND
Meiss <i>et al</i> (1993) ²⁶	Holland	ND	0/12	0/12	0/12
Ranki <i>et al</i> (1994) ²⁷	Finland	ND	1/7	ND	0/7
Alonso-Llamazares <i>et al</i> (1997) ²⁸	Spain	ND	1/9	0/6	0/6
Trevisan <i>et al</i> (1996) ²⁹	Italy	ND	3/10	ND	6/10
Buechner <i>et al</i> (1993) ³⁰	Switzerland	ND	10/10§	ND	ND
Ozkan <i>et al</i> (2000) ³¹	Turkey	ND	ND	ND	2/5
Ross <i>et al</i> (1990) ³²	Puerto Rico	10/25 (silver stain)	ND	ND	ND
Fujiwara <i>et al</i> (1997) [‡]	Japan	ND	ND	ND	2/5
Fujiwara <i>et al</i> (1997) [‡]	USA	ND	ND	ND	0/10
Daoud <i>et al</i> (1994) ³³	USA	0/10 (silver stain)	0/3	ND	ND
Fan <i>et al</i> (1995) ³⁴	USA	ND	ND	ND	0/30
Dillon <i>et al</i> (1995) ³⁵	USA	ND	ND	ND	0/20
De Vito <i>et al</i> (1996) ³⁶	USA	ND	ND	ND	0/28
Colome-Grimmer <i>et al</i> (1997) ³⁷	USA	ND	ND	ND	0/10

*Eleven of 39 cases in this study also showed an increased *B burgdorferi* induced proliferation of peripheral blood lymphocytes. †This represents the only positive culture where spirochaetes have been subtyped; in this instance PCR analysis confirmed *B afzelii*. ‡This study investigated patients from Germany, Japan, and USA; the results of each location are presented separately for clarity. §All patients in this study were selected on the basis of positive serology for borrelia infection; 7 of 7 tested also showed an increased *B burgdorferi* induced proliferation of peripheral blood lymphocytes. ND, not done; PCR, polymerase chain reaction.

Literature review

Table 1 summarises the results of the literature review.^{14–37}

DISCUSSION

The suspicion that morphoea may occur as a consequence of infection with *B burgdorferi* was initially aroused by clinical observations of coexisting morphoea and ACA, a cutaneous manifestation of chronic borreliosis.^{38–40} Subsequently, several different methodologies have been used to investigate this relation, often with confusing and conflicting results (table 1). Initially, studies used positive serology for *B burgdorferi* as evidence of previous infection and sought to correlate this with clinical manifestations of morphoea. In several series, all patients with morphoea tested have been seronegative, including 53 patients included in two studies from the UK.^{8,9,21,25,26,33} Conversely, other studies have found specific antibodies to *B burgdorferi* in between 6% and 54% of unselected patients with morphoea.^{14–18,22,23,27–29} In addition to a humoral immune response to the organism, large numbers of patients with morphoea may display specific cellular immunoreactivity.^{18,30}

Because negative serology does not exclude previous infection with *B burgdorferi* and because positive serology may merely represent coincidental infection, other studies sought more definite evidence of a causal link by seeking to demonstrate the organism in biopsies of lesional skin taken from patients suffering from morphoea. Attempts to visualise borrelia organisms directly in histological sections after silver staining or immunohistochemistry have met with mixed success, and in only a small number of cases have spirochaetes been demonstrated.^{14,15,17,19,20,25,32,33} In view of problems in evaluating positive results using these methodologies, recent

studies have focused on more sensitive and specific PCR techniques to demonstrate the organism. Once more the results have been contradictory. Studies reporting a positive association between *B burgdorferi* infection and morphoea have shown evidence of the organism in between 26% and 100% of cases,^{7,21,29,31} whereas in a further 10 reports, including our current one, no positive cases have been identified in a total of 190 cases tested.^{22,23,26–28,34–37} Lastly, the culture of borrelia from biopsies of morphoeic lesions has also been attempted. Although several studies have produced completely negative results,^{9,19,25,26,28} success has been achieved in a small number of cases,^{14,16,17,20,24} and the negative results may be partly or wholly attributable to the fastidiousness of the organism in culture.

Two possible conclusions can be drawn from this diaspora of results. The less charitable argument is that all of the positive findings reported are the result of one or more of the following: clinical and/or histological misdiagnosis of ACA as morphoea, because both entities share several common features; the chance occurrence of *B burgdorferi* infection and morphoea, especially in cases from areas of endemic Lyme disease; the presence of another causative agent that may be transmitted at the same time as *B burgdorferi*; and contamination of samples, especially with regard to PCR studies. However, it is difficult to envisage that all positive results to date can be explained in this way. This is particularly so for one group of Austrian researchers who have consistently found positive titres for *B burgdorferi* in a significantly higher proportion of patients with morphoea than in controls, and who have successfully demonstrated *B burgdorferi* in biopsies of morphoeic skin by immunoperoxidase with specific antibodies, in addition to the use of culture and typing.^{14–20} If the positive results are taken at face value then an alternative

explanation, as previously proposed, is that subspecies variations in *B burgdorferi*, which occur between different geographical locations, dictate the clinical manifestations that follow infection, with only certain strains possessing the characteristics required to initiate the development of morphoea.⁷ This phenomenon is already well documented for other accepted facets of borrelia infection. For example, ACA rarely occurs during the course of Lyme disease in North America where *B burgdorferi sensu stricto* is the dominant species, but is commonly seen in Europe where *B afzelii* and *B garinii* are more prevalent.⁴¹

"These results indicate that in the Scottish Highlands there is no association between infection with indigenous strains of *Borrelia burgdorferi* and the subsequent development of morphoea"

If this hypothesis is correct, then the literature suggests that *B burgdorferi* is implicated in the pathogenesis of at least some cases of morphoea in Austria (especially in the vicinity of Vienna),^{14–20} but almost certainly not in the USA or some, but possibly not all, parts of Germany.^{7 21–24 33–37} Isolated studies reporting a positive association in countries such as Italy, Switzerland, Puerto Rico, Turkey, and Japan, and a negative association in Spain, Finland, Holland, and France^{7 19 25–32} require corroboration before definite conclusions can be drawn about these geographical locations.

The results of our study can be assessed in a similar light using the Austrian studies as reference for an example of a population in which *B burgdorferi* plays a role in the pathogenesis of morphoea. Given the cohort of patients studied, a similar positive result should have been forthcoming if the strains of borrelia endemic in the Scottish Highlands also possessed the potential to initiate the development of morphoea, because this region has the highest incidence of Lyme disease in Scotland, and probably the UK.⁴² The breakdown of morphoea subtypes studied was similar to that described in one of the Austrian papers in which a positive association was also demonstrated, meaning that case selection was unlikely to have biased our results.¹⁶ In addition, we are confident that although PCR is prone to false negative results, our technique is sufficiently sensitive to have detected the organism were it present. Positive controls were amplified in each reaction and we have also used this technique to identify *B burgdorferi* in archival material from patients with PCBC. In addition, our cases were not biased towards late stage morphoea and, coupled with the fact that *B burgdorferi* has been identified in biopsies of patients with morphoea up to 20 years after the onset of cutaneous lesions, this means that the age of the lesions sampled was unlikely to have produced a false negative result.^{14 21} Moreover, although not directly questioned, there was no evidence that our patients were receiving antibiotic treatment before biopsy and none had a history documenting previous manifestations of Lyme disease. These results indicate that in the Scottish Highlands there is no association between infection with indigenous strains of *B burgdorferi* and the subsequent development of morphoea.

Whether or not this negative result is entirely attributable to the particular strains of *B burgdorferi* encountered in the highlands, as compared with Austria, remains to be determined. However, the data currently available do not disprove this theory. We have recently typed 12 isolates of *B burgdorferi sensu lato* grown from highland ticks, five as *B afzelii* and seven as *B burgdorferi sensu stricto*, and discovered two different strains of the first organism and four of the second.⁴³ Of particular interest to our current study was the finding that all strain types differed from reference strains derived from mainland Europe. In fact, such genomic heterogeneity appears to be normal, both within and between geographical areas.^{44–46} In view of this, and considering the expanding spec-

Take home messages

- None of the 16 patients with morphoea had clinical evidence of previous infection with *Borrelia burgdorferi*
- All of the 14 cases tested were negative for *B burgdorferi* specific DNA (using the polymerase chain reaction), despite successful amplification of appropriate positive controls in every test
- Because the literature suggests that there is a strong geographical relation between *B burgdorferi* and morphoea, these results suggest that morphoea is probably not associated with the subspecies of *B burgdorferi* found in the Highlands of Scotland

trum of disease attributed to *B burgdorferi*, further studies are warranted to investigate the effect of strain type on the clinical manifestations of infection.

ACKNOWLEDGEMENTS

This work was part funded by a grant from Raigmore Hospital, Research and Endowments Fund and part funded by grant K/MRS/50/C2774 awarded by the Chief Scientist Office, Edinburgh. The authors would also like to thank Dr J McPhie, Head of Pathology, Raigmore Hospital, Inverness, for his support and I Christie for technical assistance.

Authors' affiliations

J R Goodlad, Billington R, Department of Pathology, Highland Acute Hospitals NHS Trust, Raigmore Hospital, Inverness IV2 3UJ, UK
M M Davidson, D O Ho-Yen, Department of Microbiology, Highland Acute Hospitals NHS Trust
P Gordon, Department of Dermatology, Highland Acute Hospitals NHS Trust

REFERENCES

- 1 Ashbrink E, Hovmark H. Successful cultivation of spirochetes from skin lesions of patients with erythema chronicum migrans Afzelius and acrodermatitis chronica atrophicans. *Acta Pathol Microbiol Immunol Scand [B]* 1985;**93**:161–3.
- 2 Hovmark A, Ashbrink E, Olsson I. The spirochetal etiology of lymphadenitis benigna cutis solitaria. *Acta Derm Venereol* 1986;**66**:479–84.
- 3 Burgdorfer W, Barbour AG, Hayes SF, et al. Lyme disease—a tick-borne spirochetosis? *Science* 1982;**216**:1317–19.
- 4 Mitchell PD, Reed KD, Vandermause MT, et al. Isolation of *Borrelia burgdorferi* from skin biopsy specimens of patients with erythema migrans. *Am J Clin Pathol* 1993;**99**:104–7.
- 5 Goodlad JR, Davidson MM, Hollowood K, et al. Primary cutaneous B-cell lymphoma and *Borrelia burgdorferi* infection in patients from the Highlands of Scotland. *Am J Surg Pathol* 2000;**24**:1279–85.
- 6 Weide B, Walz T, Garbe C. Is morphoea caused by *Borrelia burgdorferi*? A review. *Br J Dermatol* 2000;**142**:636–44.
- 7 Fujiwara H, Fujiwara K, Hashimoto K, et al. Detection of *Borrelia burgdorferi* DNA (*B. garinii* or *B. afzelii*) in morphoea and lichen sclerosus et atrophicus tissues of German and Japanese but not of US patients. *Arch Dermatol* 1997;**133**:41–4.
- 8 Muhlemann MF, Wright DJM, Black C. Serology of Lyme disease. *Lancet* 1986;**1**:553–4.
- 9 Lupoli S, Cutler SJ, Stephens CO, et al. Lyme disease and localized scleroderma—no evidence for a common aetiology. *Br J Rheumatol* 1991;**30**:154–6.
- 10 Steere AC, Taylor E, McHugh GL, et al. The overdiagnosis of Lyme disease. *JAMA* 1993;**269**:1812–16.
- 11 Jaworsky C. Connective tissue diseases. In: Elder D, Elenitsas R, Jaworsky C, et al, eds. *Lever's histopathology of the skin*, 8th ed. New York: Lippincott-Raven, 1997:253–85.
- 12 Davidson MM, Evans R, Ling CL, et al. Isolation of *Borrelia burgdorferi* from ticks in the Highlands of Scotland. *J Med Microbiol* 1999;**48**:59–65.
- 13 Petersen LS, Nelson AM, Su WP. Classification of morphoea (localized scleroderma). *Mayo Clin Proc* 1995;**70**:1068–76.
- 14 Aberer E, Stanek G, Er HM, et al. Evidence for spirochetal origin of circumscribed scleroderma (morphoea). *Acta Derm Venereol* 1987;**67**:225–31.
- 15 Aberer E, Kollegger H, Kristoferitsch W, et al. Neuroborreliosis in morphoea and lichen sclerosus et atrophicus. *J Am Acad Dermatol* 1988;**19**:820–5.
- 16 Aberer E, Klade H, Stanek G, et al. *Borrelia burgdorferi* and different types of morphoea. *Dermatologica* 1991;**182**:145–54.

- 17 **Aberer E**, Stanek G. Histological evidence for spirochetal origin of morphea and lichen sclerosus et atrophicus. *Am J Dermatopathol* 1987;**9**:374–9.
- 18 **Breier F**, Klade H, Stanek G, et al. Lymphoproliferative responses to *Borrelia burgdorferi* in circumscribed scleroderma. *Br J Dermatol* 1996;**134**:285–91.
- 19 **Aberer E**, Kerstan A, Klade H, et al. Heterogeneity of *Borrelia burgdorferi* in the skin. *Am J Dermatopathol* 1996;**18**:571–9.
- 20 **Breier FH**, Aberer E, Stanek G, et al. Isolation of *Borrelia afzelii* from circumscribed scleroderma. *Br J Dermatol* 1999;**140**:925–30.
- 21 **Schempp C**, Bocklage H, Lange R, et al. Further evidence for *Borrelia burgdorferi* infection in morphea and lichen sclerosus et atrophicus confirmed by DNA amplification. *J Invest Dermatol* 1993;**100**:717–20.
- 22 **Wienecke R**, Schlupen EM, Zochling N, et al. No evidence for *Borrelia burgdorferi*-specific DNA in lesions of localized scleroderma. *J Invest Dermatol* 1995;**104**:23–6.
- 23 **Weide B**, Schttek B, Klyszcz T, et al. Morphea is neither associated with features of *Borrelia burgdorferi* infection, nor is this agent detectable in lesional skin by polymerase chain reaction. *Br J Dermatol* 2000;**143**:780–5.
- 24 **Weber K**, Prac-Mursic V, Reimers CD. Spirochaetes isolated from two patients with morphea. *Infection* 1988;**16**:25–6.
- 25 **Raguin G**, Boissic S, Souteyrand, et al. No evidence for a spirochetal origin of localized scleroderma. *Br J Dermatol* 1992;**127**:218–20.
- 26 **Meiss JF**, Koopman R, van Bergen B, et al. No evidence for a relationship between *Borrelia burgdorferi* infection and old lesions of localized scleroderma. *Arch Dermatol* 1993;**129**:386–7.
- 27 **Ranki A**, Aavik E, Peterson P, et al. Successful amplification of DNA specific for Finnish *Borrelia burgdorferi* isolates in erythema chronicum migrans but not in scleroderma lesions. *J Invest Dermatol* 1994;**102**:339–45.
- 28 **Alonso-Llamazares J**, Persing DH, Anda P, et al. No evidence for *Borrelia burgdorferi* infection in lesions of morphea and lichen sclerosus et atrophicus in Spain. A prospective study and literature review. *Acta Derm Venereol* 1997;**77**:299–304.
- 29 **Trevisan G**, Stinco G, Nobile C, et al. Detection of *Borrelia burgdorferi* in skin biopsies from patients with morphea by polymerase chain reaction. *J Eur Acad Dermatol Venereol* 1996;**6**:15–19.
- 30 **Buechner SA**, Winklemann RK, Lautenschlager S, et al. Localized scleroderma associated with *Borrelia burgdorferi* infection. Clinical, histologic and immunohistochemical observations. *J Am Acad Dermatol* 1993;**29**:190–6.
- 31 **Ozkan S**, Atabay N, Fetil E, et al. Evidence for *Borrelia burgdorferi* in morphea and lichen sclerosus. *Int J Dermatol* 2000;**39**:278–83.
- 32 **Ross SA**, Sanchez JL, Taboas JO. Spirochetal forms in the dermal lesions of morphea and lichen sclerosus et atrophicus. *Am J Dermatopathol* 1990;**12**:357–62.
- 33 **Daoud MS**, Su W, Leiferman KM, et al. Bullous morphea: clinical, pathologic, and immunopathologic evaluation of thirteen cases. *J Am Acad Dermatol* 1994;**30**:937–43.
- 34 **Fan W**, Leonardi CL, Penneys NS. Absence of *Borrelia burgdorferi* in patients with localized scleroderma (morphea). *J Am Acad Dermatol* 1995;**33**:682–4.
- 35 **Dillon WI**, Saed GM, Fivenson DP. *Borrelia burgdorferi* DNA is undetectable by polymerase chain reaction in skin lesions of morphea, scleroderma, or lichen sclerosus et atrophicus of patients from North America. *J Am Acad Dermatol* 1995;**33**:617–20.
- 36 **De Vito JR**, Merogi AJ, Vo T, et al. Role of *Borrelia burgdorferi* in the pathogenesis of morphea/scleroderma and lichen sclerosus et atrophicus: a PCR study of thirty-five cases. *J Cutan Pathol* 1996;**23**:350–8.
- 37 **Colome-Grimmer MI**, Payne DA, Tying SK, et al. *Borrelia burgdorferi* DNA and *Borrelia hermsii* DNA are not associated with morphea or lichen sclerosus et atrophicus in the southwestern United States [letter]. *Arch Dermatol* 1997;**133**:1174.
- 38 **Abele DC**, Bedingfield RB, Chandler FW, et al. Progressive facial hemiatrophy (Parry-Romberg syndrome) and borreliosis. *J Am Acad Dermatol* 1990;**22**:531–3.
- 39 **Asbrink E**, Hovmark A, Olsen I. Clinical manifestations of acrodermatitis chronica atrophicans in 50 Swedish patients. *Zentralblatt für Bakteriologie Mikrobiologie und Hygiene/Reihe A* 1986;**263**:253–61.
- 40 **Asbrink E**, Brehmer-Anderson E, Hovmark A. Acrodermatitis chronica atrophicans—a spirochaetosis. A clinical and histological picture based on 32 patients: course and relationship to erythema chronicum migrans Afzelius. *Am J Dermatopathol* 1986;**8**:209–19.
- 41 **Nadelman RB**, Wormser GP. Lyme borreliosis. *Lancet* 1998;**352**:557–65.
- 42 **Ho-Yen DO**, Davidson MM, Joss AWL. How prevalent is Lyme disease in Scotland? *SCIEH Weekly Report* 1999;**33**:214–15.
- 43 **Ling CL**, Joss AWL, Davidson MM, et al. Identification of different *Borrelia burgdorferi* genomic groups from Scottish ticks. *Mol Pathol* 2000;**53**:94–8.
- 44 **Wang G**, van Dam AP, Spanjaard L, et al. Molecular typing of *Borrelia burgdorferi* sensu lato by random amplified polymorphic DNA fingerprinting analysis. *J Clin Microbiol* 1998;**36**:768–77.
- 45 **Mathiesen DA**, Oliver JH, Jr, Kalbert CP, et al. Genetic heterogeneity of *Borrelia burgdorferi* in the United States. *J Infect Dis* 1997;**175**:98–107.
- 46 **Junttila J**, Peltomaa M, Soini H, et al. Prevalence of *Borrelia burgdorferi* in *Ixodes ricinus* ticks in urban recreational areas of Helsinki. *J Clin Microbiol* 1999;**37**:1361–5.

New JCP online submission and review system

We are pleased to inform authors and reviewers of the new online submission and review system at JCP. Developed by High-Wire Press (CA, USA), Bench Press is a fully integrated electronic system that utilises the web to allow rapid and efficient submission of manuscripts. It also allows the peer review process to be conducted entirely online. We are one of the first journals in the BMJ Special Journals group to go online in this way. The aim, apart from saving trees, is to speed up the often frustratingly slow process (for both authors and editors) from submission to publication. Many reviewers might appreciate this too. Authors may submit their manuscript in any standard word processing software. Acceptable standard graphic formats include: jpeg, tiff, gif, and eps. The text and graphic files are automatically converted to PDF for ease of distribution and reviewing purposes. Authors are asked to approve their submission before it formally enters the reviewing process. On approval by the authors, the submission is passed to the editor and/or reviewers via the web. All transactions are secure.

To access the system click on "SUBMIT YOUR MANUSCRIPT HERE" on the JCP homepage: HYPERLINK <http://www.jclinpath.com>, or you can access Bench Press directly at HYPERLINK <http://submit-jcp.bmjournals.com>.

We are very excited with this new development and would encourage authors and reviewers to use the online system whenever possible. As editors, we will use it all the time, the up side being lack of need to travel to the editorial office to deal with papers, the down side having no more excuses to postpone decisions on papers because we are "at a meeting"!

The system is very easy to use and should be a big improvement on the current peer review process. Full instructions can be found on Bench Press <http://submit-jcp.bmjournals.com> and JCP online at <http://www.jclinpath.com>. Please contact Natalie Davies, Project Manager, HYPERLINK <mailto:ndavies@bmjgroup.com> for any further information.

H Holzel, P van Diest