Molecular heterogeneity of meningioma with INI1 mutation

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**Background:** INI1 (hSNF5) mutations are linked to rhabdoid tumours, but mutations in meningiomas with hot spot mutations in position 377 have also been reported.

Aims: To analyse the INI1 gene in meningioma.

Methods: Exons 1, 4, 5, and 9 of the INI1 gene were analysed by the polymerase chain reaction and direct sequencing in 80 meningiomas. For all cases, western blotting of the INI1 protein was performed.

Results: Only one of the 80 samples showed a cytosine insertion in codon 376. This mutation changed the open reading frame in almost the whole exon 9 and resulted in a longer hSNF5 protein. Complex analysis of the above described tumour sample by western blotting, DNA sequencing, and loss of heterozygosity (LOH) analysis showed that this particular meningioma consisted of heterogeneic cellular components. One of these components had a mutated INI1 gene, whereas in the other component INI1 was intact.

Conclusions: INI1 mutation is a rare event in the molecular pathology of meningiomas. It is possible for the INI1 gene to be mutated in only a proportion of meningioma cells.

**M**utations of INI1 (integrase interactor 1) have been frequently described in rhabdoid tumours, medulloblastomas and, occasionally, in lymphoid malignancies, although the search for INI1 mutations in meningiomas has been unsuccessful (lung carcinoma, sarcomas, and breast cancer). INI1 is localised on chromosome 22q and it had been analysed in tumours with loss of heterozygosity (LOH) at 22q. Allelic loss on 22q is also characteristic for meningiomas. In most cases, LOH at 22q is linked to NF2 (neurofibromin 2) mutations. This observation led many to speculate that NF2 is not the only tumour suppressor gene localised on 22q. Schmitz and co-workers have suggested that because the INI1 gene is found on 22q, it is a putative tumour suppressor involved in the pathogenesis of meningioma. They found INI1 mutations in 3% of meningiomas (four of 126 tumours). Interestingly, INI1 mutations in all four cases were in the same position: nucleotide 377 (Arg to His substitution). This intriguing finding prompted us to check the status of INI1 hot spots—exons 1, 4, 5, and 9—in 80 cases of meningioma.

**MATERIAL AND METHODS**

**Purification of the DNA templates**

Samples of tumour and corresponding DNA from peripheral blood leucocytes used in our study were obtained from patients who had undergone surgical resection in the department of neurosurgery, Polish Mother’s Memorial Hospital, Lodz, Poland. All tumours were examined histopathologically and classified according to the World Health Organisation classification of tumours of the central nervous system. DNA was isolated by means of chloroform phenol extraction.

**PCR conditions**

The polymerase chain reaction (PCR) was performed under standard conditions, using dNTPs (Perkin Elmer, Boston, Massachusetts, USA), Taq DNA polymerase (Promega, Madison, Wisconsin, USA), and a Biometra UNO Thermoblock (Biometra, Gottingen, Germany).

**Primer sequences**

The following primers were used for PCR analysis:

- Exon 1 forward primer, 5′-ATT TCG CCT TCC GGC TTC GGT-3′.
- Exon 1 reverse primer, 5′-ATG AAT GGA GAC GCG CGC TCT-3′.
- Exon 4 forward primer, 5′-GGC CAG GCC CAG CCA AGC C-3′.
- Exon 4 reverse primer, 5′-AGA ACT AAG GCG GAA TCA GC-3′.
- Exon 5 forward primer, 5′-GCT TCC ATT TCA TCT TCA GC-3′.
- Exon 5 reverse primer, 5′-GTT CCC ACG TGA GTG CAG-3′.
- Exon 9 forward primer, 5′-TGT TCC CAC CCC TAC ACT TG-3′.
- Exon 9 reverse primer, 5′-ATG AAT GGA GAC GCG CGC TCT-3′.

**Sequencing**

Sequencing was performed by means of an automatic sequencer from LiCor (Lincoln, Nebraska, USA). DNA was amplified as described above. PCR products were precipitated with glycogen. The sequencing reaction was performed by means of the Epicentre (Madison, Wisconsin, USA) sequencing kit in accordance with the manufacturer’s protocol. Products of the sequencing reaction were visualised and analysed with the LiCor automatic sequencer.

**Sequencing primers**

The following sequencing primers were used:

- Exon 1, 5′-CTG ATC CCT CGT CGC AGC CGG G-3′.
- Exon 4, 5′-CAC GGA GGG TGA GTG CAG-3′.
- Exon 5, 5′-GGC CAG GGC CAG CCA AGC G-3′.
- Exon 9, 5′-TCT GAG ATG CTC GTG GGG AG-3′.

**LOH analysis**

LOH analysis was performed only for the meningioma with the INI1 mutation. The D22S1163, D22S298, D22S257, D22S303, and D22S268 markers overlapping the hSNF5 gene...

**Abbreviations:** LOH, loss of heterozygosity; NES, nuclear export signal; NF2, neurofibromin 2; INI1, integrase interactor 1; PCR, polymerase chain reaction
Proper amino acids and nucleotide sequence in exon 9 of INI1

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CGG  ATG  AGG  CGT  CTT  GCC  AAC  ACG  GCC  CGG  GCC  TGG  TAA
R    M    R    L    A    N    T    A    P    A    W    Stop
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Changed amino acids and nucleotide sequence in exon 9 of INI1

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CGG  ATG  (insG)AG  GCG  TCT  TGC  CAA  CAC  GCC  CCC  GGC  CTG  GTA  A
R    M    E    A    S    C    Q    H    G    P    G    L    V_i
```

ins G: guanine insertion in codon 376
TA A: disrupted stop codon
V_i: first of the additional 59 amino acids

Sequence of the additional 59 amino acids

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V_i TSPSASHGH GSKQKGPPL LHLLARTEEA OQPESAILLQ GWGGWOFQV ALPGTHSIC
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The G insertion caused the change of the open reading frame, removal of the natural stop codon, and thus elongation of the protein by the additional 59 amino acids

Figure 1 Molecular mechanisms of INI1 mutation in the case of meningioma described here.

**RESULTS AND DISCUSSION**

Only one of 80 meningioma samples showed a mutation in the INI1 gene. Histological examination of this tumour specimen revealed transitional meningioma. The INI1 mutation was different from that described before in meningiomas. The mutation changed the open reading frame of exon 9, resulting in the removal of the normal stop codon and elongation of the INI1 protein by an additional 59 amino acids (fig 1). Sequencing indicated that the mutation was heterozygotic (fig 2). However, taking into consideration the fact that heterozygotic mutations are characteristic of oncogenes rather than of tumour suppressors genes, two other explanations for the sequencing result may be offered. First, contamination of the tumour sample with normal, not neoplastic, brain tissues and, second, molecular heterogeneity of the analysed meningioma. Data base analysis of the novel 59 amino acid sequence showed no significant homology with a known protein, making the hypothesis that the observed mutation causes gain of function untenable. In addition, LOH analysis performed for chromosome 22q in the mutated meningioma showed LOH for a chromosomal region overlapping hSNF5 (SWI/SNF related, matrix associated actin dependent regulator of nucleosome remodeling) gene.

**Western Blotting**

Protein samples were obtained from frozen tumour tissues. Samples were resolved on 12% sodium dodecyl sulfate polyacrylamide gels, transferred to PVDF membranes, and reacted with the anti-INI1 antibody N-14 (sc9749) C-18 (sc9751) (Santa Cruz Laboratories, Santa Cruz, California, USA) at a 1/1000 dilution. In addition, after stripping, an actin specific antibody (actin C-2 sc8432; Santa Cruz) at a 1/1000 dilution was used to visualise potential differences in protein concentrations and protein degradation.

Figure 2 Cytosine insertion in codon 376 causes the appearance of double bands, starting from the point of mutation.

Figure 3 Western blotting analysis of several meningiomas. Anti-INI1 antibodies raised against the N-terminal end of the INI1 protein were used. Lanes 1–7, meningioma cases. Lane 5, meningioma with mutation, which produces a longer INI1 protein. Lane 8, molecular weight marker.
This positive LOH result eliminated the possibility of contamination with normal tissue. Thus, both the sequencing and LOH results could be interpreted only as a consequence of molecular heterogeneity of the meningioma sample. Western blotting analysis of the tumour sample with the INI1 mutation showed that it had a longer protein product compared with other meningiomas. This supported the notion that mutated INI1 was present in less than 50% of the meningioma cells (fig 3). Molecular heterogeneity of tumours is a frequent phenomenon.

The first two authors contributed equally to this paper.

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REFERENCES


